FLORISTIC COMPOSITION AND STRUCTURAL ANALYSIS OF FLORA IN NILGIRIS BIOSPHERE RESERVE, WESTERN GHATS OF SOUTHERN INDIA

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The field survey was conducted from April 2021–December 2021 in Mettupalyam forest range of Coimbatore forest division which is an integral part of Nilgiris Biosphere Reserve. A systematic random sampling method with 60 sample plots of 20×20 m and 8 transect line survey at 0.2% sampling intensity were laid out in the study area which recorded 54 trees, 25 shrubs, 22 herbs and 22 grasses. The result revealed that the maximum dominance was observed in *Atlantia monophylla* (32.20), *Lantana camara* (43.02), *Abutilon indicum* (54.87) and *Oplismenus burmanii* (19.34%) and minimum in *Glycosmis pentaphylla* (12.32), *Euphorbia heterophylla* (2.18), *Mimosa pudica* (2.82) and *Melinis repens* (0.18%) in tree, shrubs, herbs and grass layer, respectively. Furthermore, the highest Shannon D index (H = 3.08) observed in tree layer, followed by grass layer (H = 2.76), herb layer (H = 2.28) and shrub later (H = 2.25) along with Simpson index also higher in grass layer (1-D = 0.92), tree layer (1-D = 0.91), herb layer (1-D = 0.86) and shrub layer (1-D = 0.84), respectively. This study involving the cataloguing of flora in Mettupalayam Reserve Forest is the pioneer work which provided scientific and baseline information for conservation of flora in the forested area.

Keywords: Catalogue, phytosociology attributes, reserve forest, conservation

INTRODUCTION

The Niligiri Biosphere Reserve is located in south west India and north of the Palghat $(10^{\circ} 45' \text{ N latitude and } 76^{\circ} 10'-77^{\circ} 10' \text{ E}$ longitude), spreading over an area of 5600 km². The Reserve has diverse climate due to its varied reliefs and topography. Geological evidences suggest that the underlying rocks are Archaean which were about two billion years old. The mean annual rainfall varies from 600 mm in the eastern side to 2000 mm in the western side. The dry season is from January to April. Corresponding to the gradient in rainfall, the vegetation varies from southern tropical dry thorn forest in the east to moist deciduous forest in the west with dry deciduous forest in between the two forest types (Champion & Seth 1968). The Niligiri Biosphere

Reserve is one of the most studied landscapes with regards to biodiversity in India. About 20% of all angiosperm species, 15% of butterflies and 23% of all vertebrates, excluding marine species in India were found in the Niligiri Biosphere Reserve. It is also a significant region for its endemic species. Out of the 3000 known angiosperm species, 82 (2.7%) are exclusive to this landscape (Daniels 1993). India's Nilgiri Biosphere Reserve, which is one among the 25 global hotspots of biodiversity with diverse endemic fauna and flora, is facing severe man-made ecological disturbances (Desai & Baskaran 1996, Silori & Mishra 1995 (2001).

According to the Forest Survey of India Report 2021, the total forest cover of the country is 713,789 km² which is 21.71% of India's total geographical area. Tropical forest covers only 8% of the land area and yet supports a high level of biological variety with nearly half of all known species and a huge number of undiscovered species. Tropical forests are the most diverse terrestrial ecosystems, supporting a wide range of living forms and contributing significantly to world biodiversity (Dirzo & Raven 2003, Houghton 2005, Wilson 1988). The tropical forests which are ecologically highly imperative areas; nonetheless are under threat from population growth, urbanisation, deforestation, agriculture, degradation, loss of habitat, legal and illicit logging, mining, fire and climate change (May & Stumpf 2000, Singh et al. 2006, Goparaju & Jha 2010). Plant diversity encompasses the heterogeneity and wide variation of plant forms (Mudgal 1997, Shameem et al. 2010, Chowdhury et al. 2018). The vegetation patterns fluctuate from season to season in a cyclic fashion over the years in a successional manner. The fluctuations showed each species population is responding to prevailing temperature, humidity and sunlight as in tune by the vegetation (Heady 1958). The pattern of species distribution in a community is altered by the development and deterioration of plant species, topographic features and climatic differences (Watt et al. 2007, Karaköse 2019, Karaköse & Salih 2021). The necessity of measuring species richness is crucial for ensuring biodiversity protection (Rahman et al. 2011). In addition, floristic assessment of the forest is indispensable to detect the risk of extinction, arrival of invasive species and changes in plant diversity over time. Relatively wide range of studies were carried out to determine the forest floristic composition and forest structure in Western Ghats. An extensive assessment of the distribution of vascular plants in Western Ghats were conducted in Mudanthurai Tiger Reserve, Agasthyamalai Biosphere Reserve (Ganesh et al. 1996, Rao & Raghavendra 2012). The Mettupalayam Reserve Forest of Western Ghats holds significant population of fauna owing to its diverse distribution of flora but there is no scientific and systematic study on documentation of floristic composition in Mettupalayam Reserve Forest of Nilgiris Biosphere Reserve. In this context, there is an urgent requirement to generate the latest data on its floristic composition in the reserve forest. The objective of the present study focuses on determining the floristic structure and composition of the area. It is expected that the research findings will provide scientific baseline information for conservation of flora and fauna in the forest reserve.

MATERIALS AND METHODS

Study area

The field survey was conducted from April 2021 to December 2021 in the Mettupalayam Forest range of Coimbatore Forest Division (10° 37'–11° 31' N and 76° 39'–77° 5' E). Greater part of the division is situated southwardly extending in the Western Ghats with the Northwestern parts forming the lower ranges of the Nilgiris. Systematic random sampling method was conducted in the area using transect lines and sample plots in 6 areas of Mettupalayam range namely Jaccanari, Sundapatti, Nellimalai, Hulikal, Kandiyur and Kallar while their forest types were classified based on Champion and Seth (1968).

Assessment of phytosociological characters

A transect line with 2 km length was marked in the study area for flora exploration and documentation. The size and number of sample plots were determined using the species effort curve technique (Misra 1968, Daniels 1996). Based on species effort curve, sample plots or quadrats with the size of 20×20 m for trees, $10 \times$ 10 m for shrubs, 5×5 m for herbs and 1×1 m for grasses were placed and systematically surveyed using sampling intensity at 0.2%. Sample plots were laid opposite to each other direction and the distance between the sample plots were fixed as 200 m and 50 m from the transect line. A total of 80 sample plots and 8 transect lines were laid out in the study area. In each plot, the diameter at breast height and height of trees with a diameter at breast height of more or equal to 10 cm were recorded and location points were recorded using global positioning system. Based on the geo-referencing points, each the transect line and sample plots were marked on the google earth map (Figure 1). All flora in the plots were identified and recorded by local and scientific names. The grass samples were collected and identified by using hand book on some south Indian grasses (Achariyar & Mudaliyar 1921) and google lens application.

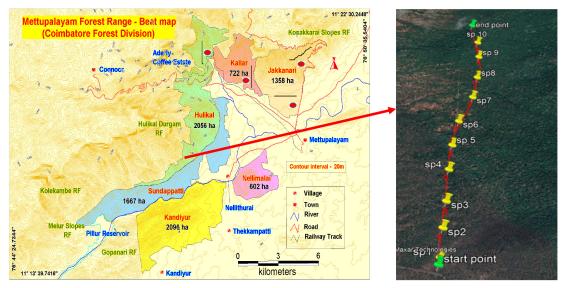


Figure 1 Map view of transect lines and sample plot locations of Mettupalyam Reserve Forest using Google Map Earth

Data analysis

Phytosociological characters

The important phytosociological characters such as density, frequency and abundance, relative

density, relative frequency, relative dominance and important value index of plant species were determined as recommended by Curtis and McIntosh (1951). The quantitative characters within the study area were assessed by using the following formula:

$$Density = \frac{Total number of individuals of a species in all quadrants}{Total number of quadrats studied}$$

$$Frequency (\%) = \frac{Number of quadrants in which the species occurred}{Total number of quadrats studied} \times 100$$

$$Abundance = \frac{Total number of individuals of a species in all quadrants}{Total number of quadrats in which the species occurred}$$

$$Relative density (\%) = \frac{Number of individual of the species}{Number of individual of all the species} \times 100$$

$$Relative frequency (\%) = \frac{Number of occurrence of the species}{Number of occurrence of all the species} \times 100$$

$$Relative dominance (\%) = \frac{Total basal area of the species}{Total basal area of all the species} \times 100$$

$$Relative Relative dominance (\%) = \frac{Total basal area of all the species}{Total basal area of all the species} \times 100$$

Species diversity indices

The following diversity indices were calculated by using PAST software (Hammer et al. 2001).

Simpson index, D =
$$\frac{\sum ni(ni-1)}{N(n-1)}$$

where D = Simpson index of dominance, ni = the total number of trees of each individual species, N = the total number of trees of all species. As the value of D increases, diversity decreases and Simpson's index was therefore usually expressed as 1 - D or 1/D (Simpson 1949).

Shannon-Weiner index, H' = – \sum pi In pi

where H' = Shannon index of diversity, pi = the proportion of important value of the ith species, pi = ni / N, ni = the important value index of it^h species and N = the important value index of all the species), ln = natural logarithm on proportion of each species (Shannon & Weiner 1963).

Pielou's evenness index, J = H/ln(S)

where H = Shannon-Weiner index, S = individuals of all the species, ln = natural logarithm on individuals of all the species (Pielou 1975).

Margalef's index, Dmg = (S-1)/ln(N)

where S = total number of species, N = total number of individuals, ln = natural logarithm on total number of individuals (Margalef 1968).

Chaol estimator,

where Q2 = number of species occurring in two samples, F1 = the number of singleton species, F2 = the number of doubleton species (Chao & Glibert 1980).

RESULTS AND DISCUSSION

Density, frequency, abundance and dominance (important value index) of flora

Tree layer

A total of 54 species were recorded which belongs to 25 families, the highest number of species was registered in Fabaceae family (Figure 2a). On the species level Atalantia monophylla (9.19) was the most dominant species whereas the lowest density was recorded in Ailanthus triphysa and Vitex altissima (0.02) (Figure 3a). With respect to frequency, Atlantia monophylla showed maximum value at 92.59%, followed by Sapindus emarginatus and Ziziphus glabrata with the value of 75.93% and minimum values were observed in ten species such as Albizia lebbeck, Ailanthus triphysa, Butea monosperma, Cassia spectabilis, Dalbergia latifolia, Diospyros montana, Glycosmis pentaphylla, Mallotus philippensis, Terminalia chebula and Vitex altissima with the frequency of 1.85% respectively (Figure 3b). Bambusa bambos had highest abundance at 12.50 followed by Atlantia monophylla at 9.92 and lowest value was recorded in Acacia planifrons at 0.66 (Figure 3c). Atalantia monophylla showed highest important value index (32.20) followed by Chloroxylon swietenia (14.38), Sapindus emarginatus (12.32) while the lowest was observed in Glycosmis pentaphylla (1.19) (Table 1). The results revealed that Atlantia monophylla had maximum dominance in Mettupalayam Reserve Forest. This is because it grows well on all type of soils and seeds dispersed by ants and birds for longer distance which enhanced the local colonisation process and plant diversity of the area (Figure 3a). The high frequency and abundance of this species were most likely due to the presence of seeds, that were easily distributed as well as their ability to regenerate quickly (Figure 3b). This study was in accordance with the findings of Onyekwelu et al. (2008), $L\ddot{u}$ et al. (2010), Zhang et al. (2012), Mandal & Joshi (2014), Sathya (2017), Abdullahi & Abba (2021) and Oluwaseyi et al. (2021) who reported the important value index in Khaya ivorensis as 14.32, Barringtonia macrostachya as 54.88, Castanopsis carlesii var. spinulosa as 9.31, Anogeissus latifolia as 37.90, Diospyros melanoxylon as 36.04, Protium serratum as 34.69, Azadirachta indica as 37.10 and Mangifera indica as 22.93. However, some values were found to be lower than the findings of Shahid & Joshi (2016) in Shorea robusta at 187, Gebrewahid & Abrehe (2019) in Acacia abyssinica as 40.85, Okechalu et al. (2021) in Daniellia oliveri as 81.90, Sahu et al. (2019) in Mangifera indica as 112.76 which compared the ecological significance of species and indicated the extent of dominance of a species in the structural system of vegetation stand.

Shrub layer

A sum of 25 shrubs were recorded in the shrub layer which belongs to 17 families and the higher number of species was observed in Euphorbiaceae family (Figure 2b). The maximum density (27.17) was recorded for *Lantana camara* followed by Solanum violaceum (14.09), Chromolaena odorata (13.37) (Figure 3a). The lowest value was registered in Calotropis procera (0.07). Jasminum angustifolium and Lantana camara had highest frequency percentage of 94.44 followed by Solanum violaceum at 79.63% and Carmona retusa showed lowest frequency percentage of 1.85 (Figure 3b). Abundance of Lantana camara exhibited maximum value of 28.76 followed by Chromolaena odorata (19.00), Solanum violaceum (17.70) compared to Euphorbia heterophylla L. (1.00) (Figure 3c) which led to increase in the important value index of Lantana camara, Solanum violaceum and Chromolaena odorata as 43.02, 26.72 and 25.03 respectively (Table 2). Lantana camara showed higher dominance in shrub layer (Figure 3a) which might be due to biological attributes such as fitness homeostasis, phenotypic plasticity, dispersal benefits from destructive foraging activities, widespread geographic range, vegetative reproduction, fire tolerance, better competitive ability than native flora and allelopathy. These are some of the biological attributes that contributed to Lantana's success as an invader species and abundant wind dispersed seeds which provided a greater reproductive capacity (Karaköse et al. 2018). Lantana camara invasions in natural communities were frequently linked to a reduction in species diversity as well as changes in ecosystem structure and function. Similar dominance of Lantana camara was also reported in moist deciduous forest of Odisha by Reddy & Pattanaik (2009), Khanduri et al. (2017) and Tiwari & Ravikumar (2018) respectively.

Herb layer

Twenty-two herb species were documented in the herb layer which belongs to 16 families and Euphorbiaceae had highest number of species (Figure 2c). The highest herb density had been

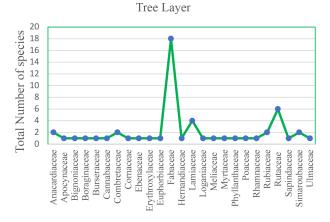


Figure 2a Distribution of family in Tree Layer

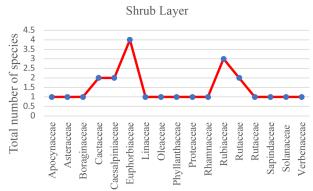


Figure 2b Distribution of family in Shrub Layer

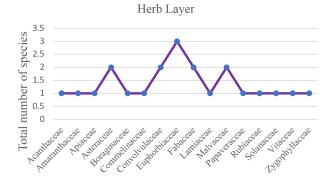


Figure 2c Distribution of family in Herb Layer

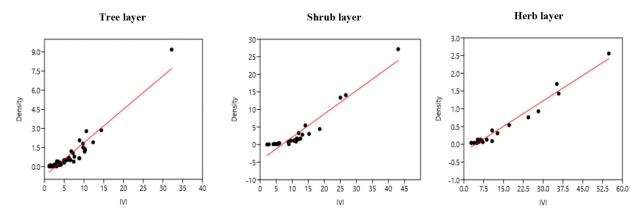


Figure 3a Density vs importance value index, *Atlantia monophyla* (Tree), *Lantana camara* (Shrub), *Abutilon indicum* (Herb) showing dominance in Mettupalyam Reserve Forest

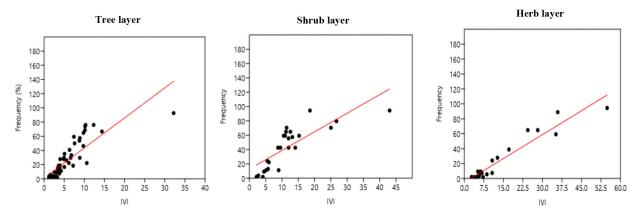


Figure 3b Frequency (%) vs importance value index, maximum frequency percentage shown by Atlantia monophylla (Tree), Jasminum angustifolium and Lantana camara (Shrub), Abutilon indicum and Barleria prionitis (Herb)

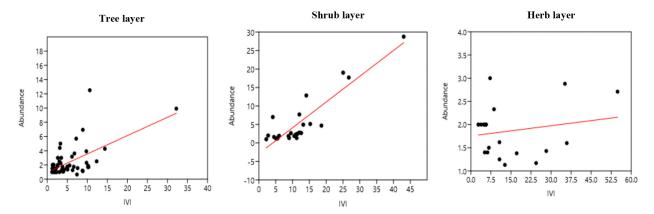


Figure 3c Abundance vs importance value index, maximum abundance shown by *Bambusa bambos* (Tree), Lantana camara and Chromolaena odorata (Shrub), Ocimum tenuiflorum, Randia tomentosa, Abutilon indicum, Solanum trilobatum, Ageratum conyzoides, Argemone mexicanna, Dichondra repens, Euphorbia hirta, Heliotropium indicum, Mimosa pudica and Tribulus terrestris (Herb). The maximum outliers observed in herb layer.

Table 1 Phytosociological attributes of trees

Sl. No	Tree	F	D	F (%)	А	IVI
1.	Acacia koenji	Fabaceae	0.11	5.56	2.00	1.56
2.	Acacia leucophloea	Fabaceae	0.09	3.70	2.50	3.01
3.	Acacia planifrons	Fabaceae	0.39	59.26	0.66	7.41
4.	Aegel marmelos	Rutaceae	0.06	3.70	1.50	1.61
5.	Aibizia lebbeck	Fabaceae	0.09	1.85	5.00	3.28
6.	Ailanthus excelsa	Simaroubaceae	0.30	16.67	1.78	5.03
7.	Ailanthus triphysa	Simaroubaceae	0.02	1.85	1.00	3.05
8.	Alangium salvifolium	Cornaceae	0.41	9.26	4.40	3.13
9.	Albizia amara	Fabaceae	1.81	46.30	3.92	9.75
10.	Atlantia monophylla	Rutaceae	9.19	92.59	9.92	32.20
11.	Azadirachta indica	Meliaceae	0.52	40.74	1.27	6.33
12.	Bahunia racemosa	Fabaceae	0.65	57.41	1.13	8.87
13.	Bamboosa bamboo	Poaceae	2.78	22.22	12.50	10.61
14.	Butea monosperma	Fabaceae	0.02	1.85	1.00	1.97
15.	Cassia fistula	Fabaceae	0.20	18.52	1.10	3.85
16.	Cassia spectabilis	Fabaceae	0.04	1.85	2.00	1.39
17.	Celtis philipensis	Cannabaceae	0.39	12.96	3.00	3.50
18.	Chloroxylon swetinea	Rutaceae	2.85	66.67	4.28	14.38
19.	Commiphora caudata	Burseraceae	0.15	11.11	1.33	4.22
20.	Cordia gharaf	Boraginaceae	0.17	9.26	1.80	2.57
21.	Dalbergia latifolia	Fabaceae	0.02	1.85	1.00	1.49
22.	Dalbergia paniculata	Fabaceae	0.26	18.52	1.40	3.99
23.	Delonix regia	Fabaceae	0.13	7.41	1.75	3.79
24.	Diospyros montana	Ebenaceae	0.04	1.85	2.00	1.31
25.	Erythroxylon monogynum	Erythroxylaceae	1.50	64.81	2.31	9.79
26.	Glycosmis pentaphylla	Rutaceae	0.02	1.85	1.00	1.19
27.	Gmelina arborea	Lamiaceae	0.50	29.63	1.69	4.96
28.	Gyrocarpus americanus	Hernandiaceae	0.50	25.93	1.93	5.58
29.	Hardwickia binata	Fabaceae	0.13	9.26	1.40	3.81
30.	Holoptelea integrifolia	Ulmaceae	0.52	29.63	1.75	6.61
31.	Limonia acidissima	Rutaceae	0.06	3.70	1.50	1.24
32.	Mallotus philippensis	Euphorbiaceae	0.06	1.85	3.00	2.62
33.	Morinda citrifolia	Rubiaceae	0.04	3.70	1.00	2.30
34.	Morinda tinctoria	Rubiaceae	0.04	3.70	1.00	2.27
35.	Muraya koenigii	Rutaceae	1.20	33.33	3.61	6.78
36.	Phyllanthus embellica	Phyllanthaceae	0.33	14.81	2.25	3.35
37.	Pongamia binata	Fabaceae	0.24	18.52	1.30	3.61
38.	Premna tomentosa	Lamiaceae	0.48	35.19	1.37	5.07
39.	Prosopis juliflora	Fabaceae	2.06	29.63	6.94	8.87
40.	Pterocarpus marsupium	Fabaceae	0.43	27.78	1.53	4.89
41.	Rhus mysorensis	Anacardiaceae.	0.30	27.78	1.07	4.00
42.	Spathodea campanulata	Bignoniaceae	0.06	3.70	1.50	2.17
43.	Sapindus emarginatus	Sapindaceae	1.91	75.93	2.51	12.32
44.	Senegalia pennata	Fabaceae	1.39	74.07	1.88	10.19
45.	Spondias mangifera	Anacardiaceae	1.17	68.52	1.70	10.16
46.	Strychnos nux vomica	Loganiaceae	0.78	50.00	1.56	7.58
47.	Syzygium cumini	Myrtaceae	0.17	14.81	1.13	3.88
48.	Tamarindus indica	Fabaceae	0.65	53.70	1.21	8.78
49.	Tectona grandis	Lamiaceae	1.06	18.52	5.70	7.22
50.	Terminalia arjuna	Combretaceae	0.70	22.22	3.17	6.17
51.	Terminalia chebula	Combretaceae	0.04	1.85	2.00	2.61
52.	Vitex altissima	Lamiaceae	0.02	1.85	1.00	1.75
53.	Wrightia tinctoria	Apocynaceae	0.07	3.70	2.00	1.62
54.	Ziziphus glabrata	Rhamnaceae	1.30	75.93	1.71	10.31

F = family, D = density, F (%) = frequency (%), A = abundance, IVI = importance value index

Sl. No	Shrub	F	D	F (%)	А	IVI
1.	Calotropis procera	Apocynaceae	0.07	3.70	2.00	2.75
2.	Carissa carandus	Apocynaceae	0.43	22.22	1.92	6.01
3.	Carmona retusa	Boraginaceae	0.13	1.85	7.00	4.14
4.	Cassia auriculata	Caesalpinioideae	0.80	42.59	1.87	8.80
5.	Catunaregam spinosa	Rubiaceae	0.20	12.96	1.57	5.74
6.	Chromolaena odorata	Asteraceae	13.37	70.37	19.00	25.03
7.	Dodonaea viscosa	Sapindaceae	1.35	59.26	2.28	10.96
8.	Euphorbia antiquorum	Euphorbiaceae	0.15	11.11	1.33	8.99
9.	Euphorbia heterophylla	Euphorbiaceae	0.02	1.85	1.00	2.18
10.	Euphorbia tirucalli	Euphorbiaceae	0.13	11.11	1.17	5.15
11.	Flueggea leucopyrus	Phyllanthaceae	0.87	64.81	1.34	11.24
12.	Grevilia hirsuta	Proteaceae	0.30	24.07	1.23	5.53
13.	Hugonia mystax	Linaceae	1.56	55.56	2.80	12.03
14.	Jasminum angustifolium	Oleaceae	4.43	94.44	4.69	18.59
15.	Jatropha gossypifolia	Euphorbiaceae	1.74	70.37	2.47	11.43
16.	Lantana camara	Verbenaceae	27.17	94.44	28.76	43.02
17.	Opuntia littoralis	Cactaceae	1.11	59.26	1.88	10.52
18.	Opuntia stricta	Cactaceae	0.15	9.26	1.60	4.50
19.	Pavetta indica	Rubiaceae	5.46	42.59	12.83	14.09
20.	Pterolobium hexapetalum	Caesalpiniaceae	3.04	59.26	5.13	15.26
21.	Randia tomentosa	Rubiaceae	1.74	64.81	2.69	12.59
22.	Solanum violaceum	Solanaceae	14.09	79.63	17.70	26.72
23.	Glycosmis pentaphylla	Rutaceae	3.28	42.59	7.70	12.03
24.	Toddalia asiatica	Rutaceae	2.83	57.41	4.94	13.19
25.	Ziziphus oenoplia	Rhamnaceae	1.13	42.59	2.65	9.52

 Table 2
 Phytosociological attributes of shrubs

F = family, D = density, F (%) = frequency (%), A = abundance, IVI = importance value index

recorded in *Abutilon indicum* (2.56) followed by Randia tomentosa (1.70), Barleria prionitis (1.43) and Sida cordifolia (0.93) which were commonly distributed whereas the lowest density was observed in Centella asiatica, Dichondra repens, Euphorbia hirta, Heliotropium indicum and Mimosa pudica with the density of 0.04 (Figure 3a). The species with high frequency in the forests was Abutilon indicum (94.44%) followed by Barleria prionitis (88.89%) and Sida cordifolia (64.81%) (Figure 3b). The high number of individuals per unit area could be due to the rapidly distributed seeds by wind and other combine factors like dormancy, germination capacity, soil moisture content and survival of fitness. Six species such as Centella asiatica, Dichondra repens, Euphorbia hirta, Heliotropium indicum, Mimosa pudica and

Ocimum tenuiflorum registered low frequency of 1.85% in the forest. Ocimum tenuiflorum had higher abundance (3.00) whereas Parthenium hysterophorus showed lower abundance (1.13) in the forests (Figure 3c). The maximum important value index of herb species was recorded in Abutilon indicum (54.87), Barleria prionitis (35.88), Randia tomentorum (35.18) and Sida cordifolia (28.20) while minimum percentage was observed in two species such as Heliotropium indicum and Mimosa pudica (2.82) (Table 3). In the herb layer, higher dominance was shown by Abutilon indicum (Figure 3a) which was due to survival, establishment and vigorous growth pattern of this species. The similar finding was observed by Mohamed and Al-Shehri (2015) for Kanawa forest.

Sl. No	Herb	F	D	F (%)	А	IVI
1.	Abutilon indicum	Malvaceae	2.56	94.44	2.71	54.87
2.	Acalypha indica	Euphorbiaceae	0.76	64.81	1.17	24.39
3.	Ageratum conyzoides	Asteraceae	0.07	3.70	2.00	5.64
4.	Argemone mexicanna	Papaveraceae	0.07	3.70	2.00	5.64
5.	Barleria prionitis	Acanthaceae	1.43	88.89	1.60	35.88
6.	Centella asiatica	Apiaceae	0.04	1.85	2.00	4.90
7.	Cissus quadrangularis	Vitaceae	0.09	7.41	1.25	10.70
8.	Commelina benghalensis	Commelinaceae	0.54	38.89	1.38	17.14
9.	Dichondra repens	Convolvulaceae	0.04	1.85	2.00	4.90
10.	Digera arvensis	Amaranthaceae	0.11	7.41	1.50	6.73
11.	Euphorbia geniculata	Euphorbiaceae	0.13	9.26	1.40	6.23
12.	Euphorbia hirta	Euphorbiaceae	0.04	1.85	2.00	3.86
13.	Heliotropium indicum	Boraginaceae	0.04	1.85	2.00	2.82
14.	Ipomoea nil	Convolvulaceae	0.39	24.07	1.62	10.70
15.	Medicago denticulata	Fabaceae	0.13	9.26	1.40	5.19
16.	Mimosa pudica	Fabaceae	0.04	1.85	2.00	2.82
17.	Ocimum tenuiflorum	Lamiaceae	0.06	1.85	3.00	7.18
18.	Parthenium hysterophorus	Asteraceae	0.31	27.78	1.13	12.72
19.	Randia dumetorum	Rubiaceae	1.70	59.26	2.88	35.18
20.	Sida cordifolia	Malvaceae	0.93	64.81	1.43	28.20
21.	Solanum trilobatum	Solanaceae	0.13	5.56	2.33	8.65
22.	Tribulus terrestris	Zygophyllaceae	0.07	3.70	2.00	5.64

 Table 3
 Phytosociological attributes of herbs

F = family, D = density, F (%) = frequency (%), A = abundance, IVI = importance value index

Grass layer

In the grass layer, *Oplismenus burmannii* showed maximum density (19.34%) followed by *Apluda mutica* (9.10%), *Chrysopogon aciculatus* (8.06%), *Cynodon dactylon* (7.43%), *Heteropogon contortus* (7.52%) whereas *Melinis repens* registered minimum density (0.18%) (Table 4). The buried grass seeds could survive in soil for long period of time and germinate under favourable climatic conditions. Moreover, these grasses spread through rhizomes and interconnected extensive root systems which contributed to their rapid growth and propagation. Consistent result was observed by Khanduri et al. (2017) for *Apluda mutica, Anaphalis busua* and *Echinochloa colona* in Garhwal, Himalaya.

Species diversity and Evenness Index

Among the four different flora habit types, tree layer registered maximum taxa of 54 followed by shrub layer at 25, grass layer at 24 and herb layer at 22. The expected species richness was similar to the observed species richness except for the tree layer at 55.67. The highest Margalef species richness, was observed in tree layer as 6.94, followed by herb layer at 3.36, grass layer at 3.16 and shrub layer at 2.84. The number of individuals more than 1000 were in tree layer, shrub layer and grass layer while it was lowest in herb layer. Species dominance was low in tree layer at 0.09 and grass layer at 0.08 as compared to herb layer at 0.14 which led to increase species diversity index. Furthermore, species diversity and similarity index were found high in tree layer and grass layer. The highest Simpson index was observed in grass layer (1-D = 0.92), tree layer (1-D = 0.91), herb layer (1-D = 0.86) and shrub layer (1-D = 0.84). The maximum Shannon D index (H = 3.08) was observed in tree layer, followed by grass layer (H = 2.76), herb layer (H = 2.28) and shrub layer (H = 2.25) respectively. The species evenness index revealed less evenness

SI. No	Grass	Density (%)
1.	Alloteropsis cimicina	5.30
2.	Apluda mutica	9.10
3.	Arachne racemose	7.05
4.	Aristida setacea	0.64
5.	Brachiaria semiundulata	3.18
6.	Bromus diandrus	0.28
7.	Bulbostylis barbata	5.53
8.	Chloris barbata	1.49
9.	Chloris virgata	0.92
10.	Chrysopogon aciculatus	8.06
11.	Cynodon dactylon	7.43
12.	Cyperus rotundus	2.95
13.	Dichanthium aristatum	5.19
14.	Digitaria ciliaris	2.50
15.	Digitaria longifolia	2.15
16.	Enteropogon monostachyus	6.20
17.	Eragrostiella bifaria	2.46
18.	Eragrostis cilianensis	0.27
19.	Heteropogon contortus	7.52
20.	Hyparrhenia hirta	0.80
21.	Melinis repens	0.18
22.	Oplismenus burmannii	19.34
23.	Perotis indica	0.71
24.	Themeda triandra	0.77

Density of grasses in the reserve forest Table 4

Table 5 Different diversity indices for Mettupalayam Reserve Forest

Diversity indices	Trees	Shrubs	Herbs	Grasses
SR	54.00	25.00	22.00	24.00
In	2071.00	4619.00	522.00	1460.00
D	0.09	0.17	0.14	0.08
1-D	0.91	0.84	0.86	0.92
Н	3.08	2.25	2.28	2.76
J	0.40	0.38	0.45	0.66
Dmg	6.94	2.84	3.36	3.16
BP	0.24	0.32	0.26	0.17
Chao-1	55.67	25.00	22.00	24.00

 $SR = observed \ species \ richness, \ ln = natural \ logarithm, \ Chao-1 = expected \ species \ richness \ to \ estimate \ total \ richness \ at \ a \ site, \ 1-D = Simpson \ index, \ H = Shannon \ index, \ J = Evenness \ index, \ Dmg = Margalef \ index, \ Shannon \ index, \ J = Evenness \ index, \ Dmg = Margalef \ index, \ Shannon \ index, \ J = Evenness \ index, \ Dmg = Margalef \ index, \ Shannon \ index, \ J = Evenness \ index, \ Shannon \ index, \ J = Simpson \ index, \ Shannon \ index, \ J = Simpson \ index, \ Shannon \ index, \ Shannon \ index, \ J = Simpson \ index, \ Shannon \ index, \ i$ BP = Berger-Parker dominance.

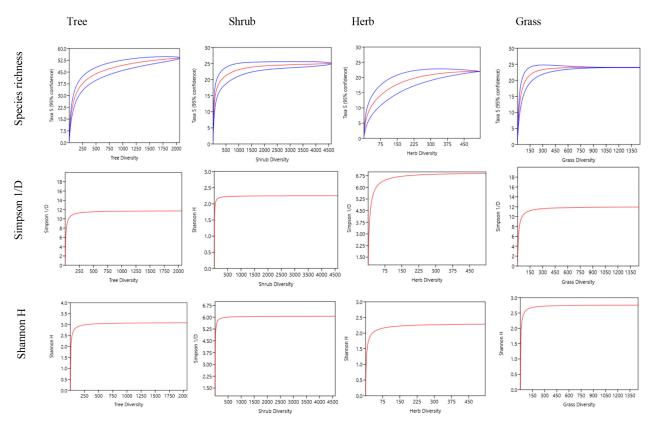


Figure 4 Species accumulation curve shows diversity index of Mettupalayam Reserve Forest

in grass layer (J = 0.66), herb layer (E = 0.45), tree layer (J = 0.40) and shrub layer (J = 0.38) which indicated the recorded species were not evenly distributed in the forest and were moderately distributed in all sites of the forest. The species richness was more in tree layer when compared to other three layers. Higher species richness may be contributed to location, soil characteristics, soil type, and climatic conditions which in turn increased the species diversity. The results were in concurrent with the findings of Timilsina et al. (2007) which observed that the number of species in tree layer was high in Sal Forest in the western Terai of Nepal.

Species accumulation curve showed diversity indices of flora in Mettupalayam Reserve Forest (Figure 4). In comparison to the other three layers, the tree layer diversity perspective may be more in terms of Shannon index of diversity values ranged from 2.25 to 3.08 (Figure 4). The present findings showed contrast with the result of Okechalu et al. (2021) which recorded the Shannon value fluctuated from 1.57 to 2.17 for Pandam Wildlife Park. The Simpson index showed a ranged from 0.84 to 0.92 and it was consistent with the work of Abdullahi (2010), Abba et al. (2015), Ogunjemite (2017), Ikyaagba et al. (2019) and Sahu et al. (2019) respectively.

CONCLUSION

Corridor management and conservation requires authenticate and consistent information on structural composition of flora and their diversity pattern. This is the pioneer research on cataloguing flora in Mettupalayam Reserve Forest. Floral composition and stand structure serving as important indicators in the formation of protection measures. This study provides scientific and baseline information on the availability of flora and its distribution pattern. Hence, it will used for management and protection of native species.

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