

## EFFECTS OF BIOFERTILISERS ON SEED GERMINATION, AND SEEDLING GROWTH AND CHEMICAL CONSTITUENTS IN NEEM (*AZADIRACHTA INDICA*)

**B. Vijayakumari**

Department of Botany, Avinashilingam University, Coimbatore 43, Tamil Nadu, India.  
E-mail: [bwiji\\_007@yahoo.co.in](mailto:bwiji_007@yahoo.co.in)

&

**K. Janardhanan**

Seed Technology Laboratory, Department of Botany, Bharathiar University, Coimbatore 46, Tamil Nadu, India

There is a growing need for utilisation of waste lands in agriculture and forestry in India. Many degraded lands and mined soils are brought to productivity, for example by using biofertilisers and organic manures. In recent years, not only in India but also in several other developing countries, biofertilisers have become an effective supplement to conventional chemical fertilisers. There is also a widening gap between the production and demand of chemical fertilisers in India and biofertilisers are able to reduce this gap and help produce high returns even from marginal and low productivity lands (Chakrabarthy 1990).

Neem (*Azadirachta indica*) belongs to the family Meliaceae. It is a tree of global importance not only due to its beautiful foliage but also for its importance in environmental conservation, now commonly introduced in agroforestry and afforestation programmes. However, there are problems in germinating neem seeds. Only fresh seeds germinate and the seeds lose viability after three months. A study was conducted to determine the effect of biofertilisers on seed germination, seedling growth and biochemical changes in neem.

Seeds collected from a 30-year-old plantation of neem were used in the study. Biofertilisers, namely, *Azospirillum*, phosphobacteria and AM fungi were obtained from the Department of Agricultural Microbiology, Tamil Nadu Agricultural University, Coimbatore. *Azospirillum* and phosphobacteria were loaded on sterilised lignite as carrier while AM fungi were loaded on vermiculite. The concentration for *Azospirillum* was  $2 \times 10^8$  and for phosphobacteria,  $5 \times 10^7$  cells  $g^{-1}$  of lignite on dry weight basis. The inoculum load of AM fungi was  $10^5$  spores  $g^{-1}$ . The soil used in the pot culture study was a typical red sandy loam with pH 7.13, E.C. 0.26 mM/dsm and organic matter 1.20%. The pots used had 9 kg capacity.

The following were the treatments studied:

- T<sub>1</sub> *Azospirillum*, 18 g/pot
- T<sub>2</sub> Phosphobacteria, 18 g/pot
- T<sub>3</sub> AM fungi, 45 g/pot
- T<sub>4</sub> *Azospirillum* + phosphobacteria, 9 + 9 g/pot respectively
- T<sub>5</sub> *Azospirillum* + AM fungi, 9 + 22.5 g/pot respectively
- T<sub>6</sub> Phosphobacteria + AM fungi, 9 + 22.5 g/pot respectively
- T<sub>7</sub> *Azospirillum* + phosphobacteria + AM fungi, 6 + 6 + 15 g/pot respectively
- T<sub>8</sub> Uninoculated as control

The biofertilisers were mixed in the soil at a rate of 2 g kg<sup>-1</sup> of soil and AM fungi at 5 g kg<sup>-1</sup> of soil. These are as per the recommendations of Tamil Nadu Agricultural University, Coimbatore.

A total of 50 seeds were sown in each pot. All pots received proper management practices and per cent seed germination was recorded a month after sowing while other growth parameters of the seedlings were recorded 120 days after sowing.

Chlorophyll content, carbohydrate level, reducing sugars, proteins, amino acids and plant phenolics were also determined 120 days after sowing. Leaf samples were estimated for chlorophyll a and b following the method of Yoshida *et al.* (1971). Total soluble carbohydrate were estimated by the anthrone reagent method (Hedge & Hofreiter 1962). The amount of total reducing sugar was estimated following the method of Somogyi (1952) while total free amino acids were estimated using the method of Moore and Stein (1948). The estimation of buffer-soluble proteins was carried out following the method of Lowry *et al.* (1951) and total phenolics were estimated by the method of Sadasivam and Manickam (1992). Data recorded were subjected to analysis of variance using completely randomised design (Panse & Sukhatme 1967).

The study revealed that seed inoculation with biofertilisers and AM fungi recorded 97% germination (Table 1). This observation is in agreement with the earlier investigations of Ponnuswamy (1993) and Vanangamudi *et al.* (1993) who reported that seed pelleting with VA mycorrhiza or phosphobacteria or *Azospirillum* significantly enhanced the germination of neem seeds. *Azospirillum*, phosphobacteria and AM fungi individually and collectively enhanced the percentage of germination by making available the nutrients, namely, phosphorus and nitrogen to help improve germination (Cooper 1979).

Inoculation with biofertilisers, namely, *Azospirillum*, phosphobacteria and AM fungi enhanced root and shoot lengths significantly compared with the rest of the treatments (Table 1). Dela Cruz *et al.* (1988) also recorded similar observations with biofertilisers. The increase in seedling biomass may be attributed to improved accumulation of nitrogen due to *Azospirillum* and of phosphorus due to AM fungi and phosphobacteria inoculation (Sanders *et al.* 1975).

Significant differences were observed in the concentrations of chlorophyll a and b due to inoculation with biofertilisers (Table 2). This may be attributed to more chlorophyll synthesis following inoculation with biofertilisers (Narayanan *et al.* 1990).

**Table 1** Effects of biofertilisers on seed germination, shoot and root length and fresh and dry weight of neem seedlings 120 days after sowing

| Treatment      | Germination after 30 days (%) | Shoot length (cm) | Root length (cm) | Fresh weight/seedling (g) | Dry weight/seedling (g) |
|----------------|-------------------------------|-------------------|------------------|---------------------------|-------------------------|
| T <sub>1</sub> | 91                            | 12.23             | 11.69            | 1.43                      | 0.66                    |
| T <sub>2</sub> | 94                            | 11.89             | 11.18            | 1.54                      | 0.67                    |
| T <sub>3</sub> | 97                            | 13.12             | 13.01            | 1.88                      | 0.86                    |
| T <sub>4</sub> | 77                            | 12.78             | 12.42            | 1.86                      | 0.74                    |
| T <sub>5</sub> | 93                            | 15.08             | 15.79            | 2.13                      | 0.96                    |
| T <sub>6</sub> | 75                            | 13.48             | 15.32            | 1.85                      | 0.83                    |
| T <sub>7</sub> | 94                            | 15.47             | 16.48            | 2.36                      | 1.09                    |
| T <sub>8</sub> | 81                            | 10.52             | 7.21             | 1.11                      | 0.42                    |
| SED            | 0.8                           | 0.05              | 0.08             | 0.05                      | 0.02                    |
| CD (p = 0.05)  | 1.6**                         | 0.10**            | 0.17**           | 1.10**                    | 0.03**                  |

SED = Standard error deviation, CD = critical difference

\*\* Significant at p < 0.01

**Table 2** Effects of biofertilisers on chlorophyll a and b, total soluble carbohydrates, reducing sugars, buffer-soluble protein, amino acid and phenolic of neem seedlings 120 days after sowing\*

| Treatment      | Chlorophyll<br>a | Chlorophyll<br>b | Total soluble<br>carbohydrate | Reducing<br>sugar | Buffer-soluble<br>protein | Amino acid | Phenolic |
|----------------|------------------|------------------|-------------------------------|-------------------|---------------------------|------------|----------|
| T <sub>1</sub> | 1.83             | 0.99             | 3.62                          | 3.69              | 37.21                     | 5.54       | 5.11     |
| T <sub>2</sub> | 1.86             | 1.00             | 4.33                          | 5.54              | 40.54                     | 3.87       | 5.68     |
| T <sub>3</sub> | 2.39             | 1.14             | 3.51                          | 5.65              | 38.50                     | 5.97       | 5.26     |
| T <sub>4</sub> | 2.13             | 1.26             | 3.21                          | 4.90              | 40.13                     | 5.24       | 4.30     |
| T <sub>5</sub> | 2.54             | 1.16             | 4.68                          | 4.57              | 53.88                     | 4.85       | 6.05     |
| T <sub>6</sub> | 2.00             | 0.94             | 3.77                          | 3.40              | 51.42                     | 5.63       | 5.08     |
| T <sub>7</sub> | 2.57             | 0.85             | 4.67                          | 7.05              | 60.83                     | 5.09       | 5.05     |
| T <sub>8</sub> | 1.61             | 0.99             | 2.78                          | 2.88              | 33.54                     | 3.23       | 3.68     |
| SED            | 0.027            | 0.02             | 0.09                          | 0.09              | 0.53                      | 0.07       | 0.06     |
| CD (p = 0.05)  | 0.054**          | 0.04**           | 0.18**                        | 0.18**            | 1.05**                    | 0.15**     | 0.12**   |

Data represent mean of three determinations

\* Expressed in mg per g of tissue on fresh weight basis

\*\* Highly significant (p < 0.01)

SED = Standard error deviation, CD = critical difference

Besides total chlorophyll, this study revealed clearly that a combined inoculation of *Azospirillum*, phosphobacteria and AM fungi significantly enhanced carbohydrates, reducing sugars, buffer-soluble proteins, amino acids and phenolics. Similar observations have been made by Niranjan *et al.* (1990) and Kanakadurga and Ramarao (1995).

From the foregoing study it is clear that the use of biofertilisers, namely, *Azospirillum*, phosphobacteria and AM fungi greatly improved seed germination and establishment and growth of neem seedlings.

## References

- CHAKRABARTY, K. P. 1990. Biofertilizers, an overview. *Everyman's Science* 25: 79–82.
- COOPER, R. 1979. Bacterial fertilizers in the Soviet Union. *Soil Fertilizer* 22: 327–333.
- DELA CRUZ, R. E., NANALO, M. Q., AGGANGAN, N. S. & AMBALO, J. D. T. 1988. Growth of three legume trees inoculated with VAM fungi and *Rhizobium*. *Plant and Soil* 108: 111–115.
- HEDGE, J. E. & HOFREITER, B. T. 1962. Determination of total carbohydrate by anthrone method. Pp. 17 in *Carbohydrate Chemistry*. Academic Press, New York.
- KANAKADURGA, V. V. & RAMARAO, P. 1995. Effect of VAM on shoot and root phenol concentration of *Terminalia arjuna*. Mycorrhizae: biofertilizers for the future. Pp. 478–481 in *Proceedings on the Third National Conference on Mycorrhiza*. 13–15 March 1995. Delhi.
- LOWRY, O. H., ROSENBROUGH, N. J., FARR, A. L. & RANDALL, R. J. 1951. Protein measurement with folin phenol reagent. *Journal of Biological Chemistry* 193: 267–275.
- MOORE, S. & STEIN, W. H. 1948. Estimation of total free amino acids. Pp. 468 in *Methods in Enzymology*. Academic Press, New York.
- NARAYANAN, R., RANGARAJAN, M. & KANDASAMY, B. 1990. Response of a Shola plant *Rhododendron nilgircum* to endomycorrhizal inoculation. Pp. 11–12 in Jalali, B. K. & Chand, H. (Eds.) *Proceedings of the National Conference on Mycorrhizae*. 14–16 February 1990. Hissar Agricultural University, Hissar.
- NIRANJAN, R., PARMILA, S. B., RAO, V. M. & SHARMA, P. 1990. Studies on the effect of rhizobium and endomycorrhizal interaction in *Dalbergia sissoo* research. Pp. 205–207 in Jalali, B. K. & Chand, H. (Eds.) *Proceedings of the National Conference on Mycorrhiza*. 14–16 February 1990. Haryana Agricultural University, Hissar.
- PANSE, V. G. & SUKHATME, P. V. 1967. *Statistical Methods for Agricultural Workers*. ICAR Publication, New Delhi.
- PONNUSWAMY, A. S. 1993. Seed Technological Studies in Neem. Ph.D. thesis, Tamil Nadu Agricultural University, Coimbatore.
- SADASIVAM, S. & MANICKAM, A. 1992. *Biochemical Methods for Agricultural Sciences*. Wiley-Eastern Ltd., New Delhi.
- SANDERS, F. E., MOSSE, B. & TINKER, P. B. 1975. *Endomycorrhizae*. Academic Press, London.
- SOMOGYI, N. 1952. Notes on sugar determination. *Journal of Biological Chemistry* 195: 19–23.
- VANANGAMUDI, K., PADMAVATHI, S., MANONMANI, V. & SURENDRAN, C. 1993. Effect of pelleting with biofertilizers, biocides and nutrients on the viability and vigour of neem seed. Pp. 64 in *World Neem Conference*. 24–28 February. Bangalore.
- YOSHIDA, S., FORNO, D. & COCK, J. H. 1971. *Laboratory Manual for Physiological Studies of Rice*. IRRI Publication, Phillipines.